

# **Test Report**



Report No.:	WD20240508

- Test Article: Poly-L-lactic acid microspheres PLLA microspheres
- Sponsor: Shenzhen Esun Industrial Co., Ltd.
- Manufacturer: Shenzhen Jusing Biotechnology Co., Ltd.
- Test Type: Commission Test
- Date of Issue: May 13,2024



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Test Article	Poly-L-lactic acid microspheres PLLA microspheres				
Test Type	Commission Test	Identification No./ Lot No.	1		
Trade Mark	esun	Model / Type	Viscosity average Molecular Weight sixty thousand		
Date of Manufacturing	Jan. 23, 2024	Accepting Date	Mar. 05, 2024		
Sponsor	Shenzhen Esun Industrial Co., Ltd.				
Applicant Address	15A, Microsoft Ketong Building, No. 55 Gaoxinnan 9th Road, Hightech Community, Yuehai Street, Nanshan District, Shenzhen.				
Manufacturer	Shenzhen Jusing Biotechnology Co., Ltd.				
Production Address	Floor 3, No. 9, Yifenghua Innovation Industrial Park, Xinshi Community, Dalang Street, Longhua district, Shenzhen City				
Test Items	In vitro Cytotoxicity Test				
Test in Accordance with	ISO 10993-5:2009 Biological evaluation of medical devices Part 5: Tests for in vitro cytotoxicity				
Summary	The test article, Poly-L-lactic acid microspheres PLLA microspheres, was extracted by MEM with 10% fetal calf serum. The extract was evaluated for in vitro cytotoxicity test by MTT assay recommended by ISO 10993-5:2009 Biological evaluation of medical devices, Part 5: Tests for in vitro cytotoxicity. L-929 cells were seeded in 96-well assay microtiter plate. After the incubation of cells for 24 h at 37°C in air with 5% CO <sub>2</sub> , blank control, negative control, positive control and four different concentrations of the test article extract were added to microtiter plate. After 24 h treatment, carefully removed the culture medium from the plates. 50µL of the MTT solution was then added to each test well and the plates were further incubated for 2 h. Then the MTT solution was discanted and 100µL of isopropanol were added in each well. Swayed this plate and subsequently transferred it to a microplate reader equipped with a 570 nm filter to read the absorbance (reference wavelength 650 nm). The reduction of viability compared to the blank (Viab%) was calculated. Under the conditions of this study, the Viab.% of the positive control group and negative control group were 16% and 100%; the Viab.% of 100%, 50%, 25% and 12.5% extracts were 96%, 99%, 99% and 102%.				
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#### **INTRODUCTION**

The test article identified below was extracted and the extract was subjected to an *in vitro* cytotoxicity test by MTT assay recommended by the Annex C of the ISO 10993-5:2009 *Biological evaluation of medical devices- Tests for in vitro cytotoxicity*.

The test article was accepted on Mar. 05, 2024. The extraction was applied from Mar. 19, 2024 to Mar. 20, 2024, and the observations were concluded on Mar. 21, 2024.

#### **MATERIALS**

The article provided by the sponsor was identified and handled as follows:			
Test article:	Poly-L-lactic acid microspheres PLLA microspheres		
Identification No./ Lot No.:			
Storage conditions:	Store at 2-8°C		
Cell line:	L929 mouse fibroblast cells, recommended by the ISO 10993-		
	5:2009, was from China Center for Type Culture Collection.		
	Cultures were incubated at 37°C in air with 5% carbon dioxide.		
MEM with phenol red:	Thermo Fisher Scientific corporation product, with Earle's salts,		
	L-Glutamine and phenol red. (Without specification, "MEM"		
	mentioned in this text mean MEM with phenol red).		
MEM without phenol red:	Thermo Fisher Scientific corporation product, with Earle's salts,		
F 1 10	without L-Glutamine and phenol red.		
Fetal calf serum:	Shanghai BasalMedia Technologies corporation product.		
Penicillin-streptomycin, liquid:	Thermo Fisher Scientific corporation product.		
MTT:	Merck corporation product.		
	MTT is solute fresh in MEM without supplements and without phenol red at a concentration of 1 mg/mL. Solution is sterilized		
	by sterile filtration using syringe filters (pore size $\leq 0.22 \ \mu$ m).		
	The solution should be used the same day.		
DMSO:	Merck corporation product.		
Isopropanol:	Guangzhou Chemical Reagent factory product.		
Extraction vehicle:	MEM with 10% fetal calf serum.(100IU/mL penicillin,		
	100µg/mL streptomycin)		
Test article preparation:	Under aseptic conditions, according to the requirements of the		
	sponsor, 1.25 g of the test article was covered with 6.25 mL		
	extraction vehicle based on a ratio of 0.2 g/mL and extracted at		
	37°C for 24 h with agitation (The extract was used		
	immediately). The extract of test article was transparent with no		
	presence of particulates. Prior to use, the extract was sterilized		
	by membrane filtration (0.22 $\mu$ m). The filtration extract of test		
	article was diluted by MEM with 10% fetal calf serum. The		
	100 %, 50%, 25% and 12.5% extracts were tested.		
Negative control preparation:	Under aseptic conditions, 2.00 g of high-density polyethylene		
	was covered with 10.00 mL extraction vehicle based on a ratio		
	of 0.2 g/mL and extracted at 37°C for 24 h. Prior to use, the		
Positive control preparation:	extract was sterilized by membrane filtration (0.22 $\mu$ m).		
Positive control preparation:	extract was sterilized by membrane filtration (0.22 $\mu$ m). MEM (1×MEM) with 10% fetal calf serum and with 10%		
Positive control preparation: Blank control preparation:	extract was sterilized by membrane filtration (0.22 $\mu$ m).		

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#### **METHODS**

**Experimental Procedure:** 

1. L-929 cell suspension preparation

- a. L-929 cell monolayer confluence was cultured with  $1 \times MEM$  for 48 h~72 h and removed from culture flasks by enzymatic digestion (trypsin/EDTA).
- b. The cells were then resuspended in culture medium and the cell suspension was adjusted at a density of  $1 \times 10^5$  cells/ml.
- 2. MTT assay
  - a. Using a multichannel pipette, dispensed 100  $\mu$ L culture medium only (blank) into the peripheral wells of a 96-well tissue culture microtitre plate. In the remaining wells, dispensed 100  $\mu$ L of a cell suspension of 1×10<sup>5</sup> cells/mL. Blank (both at the left side and the right side), negative control, positive control and the test article group were set up, and each group contained six wells.
  - b. Incubated cells for 24h (5 % CO<sub>2</sub>, 37 °C, > 90 % humidity) so that cells formed a half-confluent monolayer.
  - c. After 24 h incubation, aspirate culture medium from the cells. Per well, added 100  $\mu$ L of treatment medium containing either the appropriate concentration of test article extracts, or the negative control, or the positive control, or nothing but blank. Four different concentrations of the test item extracts were tested (100%, 50%, 25%, 12.5%).
  - d. Incubated cells for 24 h (5 %  $CO_2$ , 37°C, > 90 % humidity).
  - e. After 24 h treatment, examined each plate and cellular morphology under a phase contrast microscope. Recorded changes in the morphology of the cells due to cytotoxic effects of the test article extracts.
  - f. After the examination of the plates, carefully removed the culture medium from the plates. Pipetted 50  $\mu$ L of MTT solution into each well, and incubated the plate for 2 h at 37°C. Then the MTT solution was discanted and 100  $\mu$ L of isopropanol were added in each well. Swayed this plate and subsequently transferred it to a microplate reader equipped with a 570 nm filter to read the absorbance (reference wavelength 650 nm). To calculate the reduction of viability compared to the blank Equation was used:

$$Viab.\% = \frac{100 \times OD_{570e}}{OD_{570b}}$$

where

 $OD_{570e}$  is the mean value of the measured optical density of the extract of the test article;

 $OD_{570b}$  is the mean value of the measured optical density of the blanks.

- 3. Evaluation Criteria
  - a. A test meets the acceptance criteria if the mean  $OD_{570}$  of blanks is  $\ge 0.2$ .
  - b. A test meets acceptance criteria if the left and the right mean of the blanks do not differ by more than 15 % from the mean of all blanks.
  - c. If viability is reduced to < 70% of the blank, it has a cytotoxic potential. The 50% extract of the test sample should have at least the same or a higher viability than the 100% extract; otherwise the test should be repeated.

Referred to ISO10993-5:2009, qualitative morphological grading of cytotoxicity of extracts is given in Table 1.

Grade	Reactivity	Conditions of all cultures				
0	None	Discrete intracytoplasmatic granules, no cell lysis, no reduction of cell growth				
1	Slight	Not more than 20 % of the cells are round, loosely attached and without intracytoplasmatic granules, or show changes in morphology; occasional lysed cells are present; only slight growth inhibition observable.				

Table 1 Qualitative morphological grading of cytotoxicity of extracts

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2	Mild	Not more than 50 % of the cells are round, devoid of intracytoplasmatic granules, no extensive cell lysis; not more than 50 % growth inhibition observable.			
3	Moderate	Not more than 70 % of the cell layers contain rounded cells or are lysed; cell layers not completely destroyed, but more than 50 % growth inhibition observable.			
4	Severe	Nearly complete or complete destruction of the cell layers.			

#### **RESULTS**

The mean  $OD_{570}$  of blanks was  $\ge 0.2$  and the left and the right mean of the blanks did not differ by more than 15 % from the mean of all blanks, so the test met the acceptance criteria.

Referred to the table 1, the microscopic observation show the negative control was grade 0; the positive control was grade 4; the cytotoxicity of 100 %, 50%, 25% and 12.5% extracts were grade 1, grade 1, grade 1 and grade 0.

The results of optical density is given in Table 2:

Table 2 Absorbance and <i>Viab</i> %								
Groups	Blank Control		Positive	Negative	100%	50%	25%	12.5%
Groups	Left	Right	Control	Control	Extract	Extract	Extract	Extract
Well 1	0.9360	0.8901	0.1473	0.9321	0.8838	0.8935	0.9209	0.9449
Well 2	0.9099	0.8991	0.1224	0.8915	0.8789	0.9126	0.9255	0.9277
Well 3	0.9220	0.9174	0.1338	0.9259	0.8973	0.9089	0.9033	0.9187
Well 4	0.9461	0.9553	0.1699	0.9340	0.9269	0.9655	0.9298	0.9649
Well 5	0.9566	0.9287	0.1638	0.9477	0.9114	0.9515	0.9296	0.9648
Well 6	0.9851	0.9999	0.1876	0.9799	0.8885	0.9091	0.9523	0.9937
Mean	0.9426	0.9318	0.1541	0.9352	0.8978	0.9235	0.9269	0.9524
OD	0.9	372	0.1341 0.955	0.9332	0.09/0	0.9233	0.9209	0.9324
Viab.%	,	/	16%	100%	96%	99%	99%	102%

#### **CONCLUSION**

Under the conditions of this study, the Viab.% of the positive control group and negative control group were 16% and 100%; the Viab.% of 100%, 50%, 25% and 12.5% extracts were 96%, 99%, 99% and 102%.

Results and conclusions apply only to the test article tested. No further evaluation of these results is made by our testing center. Any extrapolation of these data to other samples is the responsibility of the sponsor. All procedures were conducted in conformance with ISO 17025.

#### **RECORD STORAGE**

All raw data pertaining to this study and a copy of the final report are to be retained in designated archive files in our testing center.

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